

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Achieving structural diversity using the perpendicular conformation of *alpha*-substituted phenylcyclopropanes to mimic the bioactive conformation of *ortho*-substituted biphenyl P4 moieties: Discovery of novel, highly potent inhibitors of Factor Xa

Jennifer X. Qiao*, Daniel L. Cheney, Richard S. Alexander, Angela M. Smallwood, Sarah R. King, Kan He, Alan R. Rendina, Joseph M. Luettgen, Robert M. Knabb, Ruth R. Wexler, Patrick Y. S. Lam

Bristol-Myers Squibb Company, Research and Development, PO Box 5400, Princeton, NJ 08543-5400, USA

ARTICLE INFO

Article history: Received 8 April 2008 Revised 21 May 2008 Accepted 22 May 2008 Available online 29 May 2008

Keywords: Alpha-substituted phenylcyclopropyl Perpendicular conformation Ortho-substituted biphenyl Factor Xa inhibitors

ABSTRACT

Ortho-substituted biphenyl moieties are widely used in drug design. We herein report a successful use of the perpendicular conformation of the *alpha*-substituted phenylcyclopropyl groups to mimic the aplanar, biologically active conformation of the *ortho*-substituted biphenyl moieties to achieve structural diversity. This is exemplified by the design and synthesis of a series of highly potent pyrazole bicyclic-based Factor Xa (FXa) inhibitors bearing *alpha*-substituted phenylcyclopropyl P4 moieties. The designed perpendicular conformation was confirmed by the X-ray structure of FXa-bound compound **2r**. The potential structural basis for the high FXa potency in the phenylcyclopropyl P4 analogs and their improved FXa inhibitory activities compared with the biphenyl P4 counterparts are discussed.

© 2008 Elsevier Ltd. All rights reserved.

Thrombotic diseases, such as myocardial infarction, stroke, unstable angina, deep vein thrombosis, and pulmonary embolism, remain the major causes of deaths and disabilities in developed countries. Existing antithrombotic therapies using either parenteral anticoagulants, such as heparins, or the oral anticoagulants, such as warfarin (coumadin®), have multiple limitations and complications. There still exists a large unmet need for safer and more efficacious oral anticoagulants. One approach to producing such anticoagulants is to inhibit thrombin generation by targeting the inhibition of coagulation Factor Xa (FXa). FXa is a key enzyme in the coagulation cascade, located at the convergence of the extrinsic and the intrinsic activation pathways. It is believed that inhibition of FXa prevents thrombus formation without compromising normal hemostasis and platelet activation because the basal thrombin level is maintained. Selective and orally active FXa inhibitors are believed to have higher efficacy, fewer bleeding risks, and a more favorable safety/efficacy ratio, compared with current treatments.^{1,2} In the recent past, both preclinical and clinical data showed the inhibition of FXa to be an effective approach for the treatment of arterial and venous thrombosis. 1,2

We have previously reported on a series of 7-oxo-4,5,6,7-tetra-hydro-1*H*-pyrazolo[3,4-*c*]pyridine (fused-pyrazole bicyclics) FXa

inhibitors containing ortho-substituted biphenyl P4 moieties, wherein the potent substituent at the ortho position was a basic group ($R'=CH_2NR^1R^2$) with a general structure **1** (Fig. 1).³ The driving force behind the present work was to further explore the structural diversity of the P4 region⁴ of the fused-pyrazole bicyclic series 1 to identify compounds with superior potency, and in vitro and in vivo profile to razaxaban.^{5a} In this paper, we first describe our use of the perpendicular conformation of alpha-substituted phenyl-cyclopropanes (Fig. 2 shows the structure of alpha-methyl substituted phenylcyclopropane) to mimic the biological conformation of ortho-substituted biphenyl P4 groups. We then present the initial P4 SAR of a series of highly potent and selective fusedpyrazole bicyclic FXa inhibitors containing such alpha-substituted phenylcyclopropyl P4 moieties with general structure 2 (Fig. 1). We next describe the X-ray structure of a representative analog, and discuss the potential structural basis for the improved FXa inhibitory activity of this series of compounds compared with the corresponding ortho-substituted biphenyl P4 counterparts.

X-ray crystallographic analysis and modeling studies of **1a** bearing an *ortho-N,N*-dimethylaminomethyl biphenyl P4 group and its related analogs revealed that the biphenyl group extends into the S4 pocket formed by the side-chains of Y99, F174, and W215 of FXa. The two phenyls are twisted, adopting a near orthogonal conformation, which according to quantum mechanical calculations^{6a-c} contains modest strain. The *ortho* substituent (R'=CH₂NR¹R²) on

^{*} Corresponding author. Tel.: +1 609 818 5298; fax: +1 609 818 6810. E-mail address: jennifer.qiao@bms.com (J.X. Qiao).

Figure 1. Overall strategy.

the biphenyl moiety is partially positioned outside of the S4 pocket in solvent. We speculate that the nitrogen atom in R' may engage in polar interactions, possibly water-mediated, with one or more of the backbone carbonyl or side-chain of E97, or the backbone carbonyl of K96. Based on the aplanar and hydrophobic feature of the biphenyl group in 1, we hypothesized that *alpha*-substituted phenylcycloalkyls might serve as rigid frameworks to direct the substituent on the quaternary carbon to the same space occupied by the *ortho* substituent of the biphenyls (such as CH₂NR¹R² in the P4 moiety of 1).

Phenylcyclopropanes exist in two limiting conformations (Fig. 2): 7 (1) the *perpendicular conformation* **P**, in which the normal vector to the phenyl ring (the vector that is perpendicular to the plane in which the phenyl ring lies) is perpendicular to the C(2)–C(3) bond of the cyclopropane ring and (2) the *bisected conformation* **B**, in which the phenyl plane bisects the three-membered ring. As shown in Figure 3, we envisioned that we could achieve structural diversity using the perpendicular conformation of *alpha*-substituted phenylcyclopropyl to mimic the biologically active conformation of the *ortho*-substituted biphenyl moiety.

Phenylcyclopropanes without *alpha*-substitution adopt a bisected conformation in the solid state.⁸ Quantum calculations

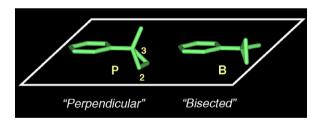


Figure 2. The perpendicular conformation (P) and the bisected conformation (B) of *alpha*-methyl phenylcyclopropane.

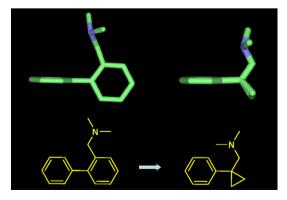


Figure 3. The bioactive conformation of 2'-CH₂NMe₂-1,1'-biphenyl group in **1a** vs the proposed perpendicular conformation of α -CH₂NMe₂phenylcyclopropyl group in **2**

and other structural analysis methods⁹ also suggested the bisected conformation to be preferable over the perpendicular conformation. Alpha-substitution on phenylcyclopropanes is expected to both sterically and electronically affect the relative conformation between the cyclopropyl and the phenyl ring.⁹⁻¹¹ A simple alphamethyl substitution resulted in the preference of the perpendicular form as indicated by both photoelectron spectra (PES)^{10a} and NMR studies. 10b In-house quantum mechanical calculations supported this observation, showing that methyl substitution at the alpha position induces distinct preferences for both the perpendicular conformation and the bisected conformation (two minima) with the perpendicular conformation marginally more favorable (0.7 kcal).¹² Surprisingly, little investigation, either experimental or theoretical, has been reported regarding the conformational preference of the alpha-substituted phenylcyclopropanes other than with simple alkyl groups.

We selected the *alpha*-substituted phenylcyclopropyl moiety as a potential P4 surrogate based on the potential conformational predisposition of the *alpha*-substituted phenylcyclopropyl ligand toward the perpendicular conformation, which mimics the biologically active conformation of *ortho*-biphenyl P4 groups, as well as based on the small size and metabolic stability of the cyclopropyl group compared with other cycloalkyls.

Table 1 depicts the initial SAR of the alpha-substituent (R group) of the phenylcyclopropyl P4 analogs. Compounds with small, neutral, and polar R groups containing one or more hydrogen bond donors or acceptors, such as COOMe (2a), CN (2b), CH2OH (2c), CH₂OMe (2d), or CONH₂ (2e), showed low nM in vitro FXa binding affinity. Though they were ten times less potent than 1a (FXa $K_i = 0.3$ nM), they were generally more potent than the corresponding biphenyl compounds with these same substituents.³ The data suggest that the cyclopropyl group, though 'wider' than the distal phenyl group, was easily accommodated by the S4 pocket of FXa. Compounds 2f-2k exemplified the variety of amide substituents we studied. Smaller alkyl amides, such as CONMe2 in 2f, as well as the heteroaryl amides, such as thiadiazole 2i and tetrazole 2k, gave subnanomolar FXa inhibition similar to that observed with the small neutral R groups in compounds 2a-2e. In contrast, compounds containing larger di-substituted amides, such as 2h and 2i, were much less active.

Replacement of the amide carbonyl in compounds 2e-2k with a methylene group resulted in significantly more potent FXa inhibitors. For instance, 2h containing a pyrrolidine amide had a FXa $K_i = 179$ nM. The corresponding des-carbonyl analog 2r had a FXa $K_i = 21$ pM, an 8000-fold increase in binding affinity. Several of the basic N-alkylaminomethyl or N,N-dialkylaminomethyl analogs achieved FXa binding affinity in the pM range. Among these, compounds 2n, 2q, 2r, 2t, and 2s, with cyclic or acyclic tertiary amines, were slightly more active than compounds with secondary amines (such as 2m and 2p), and the primary amine analog 2t. Compound 2v, bearing a less basic N-thiazolyl aminomethyl group, t0 and t1 are FXa t1 and t2 and t3 are quipotent to the more basic secondary alkyl amines such as t3 (FXa t1 and t2 pM). The basic

Table 1 a,b

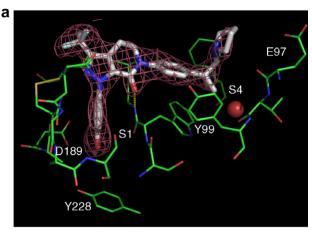
Compound	R	$FXa^b K_i (nM)$	PT ^b EC _{2x} (μM)
2a	-COOMe	1.9	83
2b	-CN	1.8	36
2c	-CH ₂ OH	2.4	nd
2d	-CH ₂ OMe	1.6	nd
2e	-CONH ₂	3.3	nd
2f	-CONMe ₂	5.8	nd
2g	-CONH-cyclopentyl	30	nd
2h	-CO-N-pyrrolidine	169	nd
2i	-CO-4-OH-piperidin-1-yl	54	nd
2j	-CONH-1,3,4-thiadiazol-2-yl	1.8	31
2k	-CONH-1H-tetrazol-5-yl	3.0	nd
21	-CH ₂ NH ₂	0.57	4.4
2m	-CH ₂ NHMe	0.18	2.9
2n	-CH ₂ NMe ₂	0.035	1.3
20	-CH ₂ NH-isopropyl	0.042	2.4
2p	-CH ₂ NH-cyclopentyl	0.12	3.2
2q	-CH ₂ NEt ₂	< 0.02	1.9
2r	-CH ₂ -N-pyrrolidine	0.021	1.4
2s	-CH ₂ -N-pyrrolidine-2-(R)-OH	0.038	2.4
2t	-CH ₂ -N-morpholine	0.064	14
2u	-CH ₂ -N-piperidine-4-OH	0.14	9.0
2v	-CH ₂ NH-thiazole-2-yl	0.054	3.4
2w	-CH ₂ -2-Me-1 <i>H</i> -imidazol-1-yl	0.025	2.7
2x	-CH ₂ -N(Me)COMe	0.055	5.6
2 y	-CH ₂ -N(Me)SO ₂ Me	0.074	24
2z	-CH ₂ -N(Me)CONHEt	0.14	12

^a All compounds were purified by either reverse phase HPLC or preparative LC/MS (water/acetonitrile or water/methanol gradient + 0.5% TFA). The basic compounds were isolated as TFA salts following lyophilization. All compounds gave satisfactory spectral and analytical data.

N-(1-methyl-imidazole-2-yl)aminomethyl analog $2\mathbf{w}^{13b}$ was also highly active with a FXa K_i of 25 pM. Unlike the *ortho-N*-methylaminomethyl biphenyl analogs previously studied, neutralization of N-methylaminomethyl moiety in $2\mathbf{m}$ to form amide $2\mathbf{x}$, sulfonamide $2\mathbf{y}$, and urea $2\mathbf{z}$ did not decrease FXa binding affinity. Because we were interested in compounds with relatively good aqueous solubility and low $\log P$, compounds with all-carbon side-chains were not studied.

The basic analogs 2l-2w, except for the morpholinyl analog 2t and the piperidinyl analog 2u, showed good anticoagulant activity with PT EC $_{2x}$ less than 5 μ M in human plasma. Despite having pM binding activity, sulfonamide 2y and urea 2z had significantly weaker anticoagulant activity compared with the parent methylamine analog 2m, presumably resulting from the increased lipophilicity. Thus, compounds 2y and 2z were projected to have higher protein binding. Compounds in Table 1 were generally selective against other related human serine proteases (inactive against trypsin, FIXa, FVIIa, plasmin, chymotrypsin, tPA, plasma kallikrein, and APC, and were more than 5000-fold selective against thrombin).

The crystal structure of $2\mathbf{r}$ (FXa K_i = 21 pM) complexed to human FXa was solved at 2.0 Å resolution with a crystallographic \mathbf{R} factor of 0.22.¹⁴ Figure 4 a depicts the electron density of $2\mathbf{r}$ in the active site of FXa. The overall L-shaped configuration is similar to that observed for other pyrazole analogs.^{3,4} The molecule adopts an extended binding mode with the para-methoxyphenyl group lo-



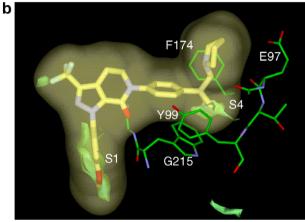


Figure 4. (a) Electron density around $2\mathbf{r}$ (FXa K_i = 21 pM) shown with the initial 2Fo-Fc electron density map contoured at 0.7σ in magenta mesh in the active site of human Factor Xa. Graphic was created using PyMol. (b) Transparent connelly surface of compound $2\mathbf{r}$ is shown in light yellow. Using the GRID program, the interaction of a hypothetical aromatic CH probe is contoured in light green at -3.5 kcal, a level that generally coincides with strong hydrophobic interactions. For simplicity, only key residues in the S4 pocket are shown.

cated in the S1 pocket and the α -CH₂-N-pyrrolidinyl phenylcyclopropyl group occupying the S4 pocket. It is noteworthy that the phenylcyclopropyl ring adopts the perpendicular conformation, which was a central feature of our inhibitor design.

Numerous in-house modeling studies and crystal structures indicated that **2r** interacts with the enzyme in essentially the same way as other pyrazole bicyclic analogs do, other than the interactions in the S4 pocket. As shown in Figure 4b, the cyclopropane ring sits above the indole ring of W215 and is sandwiched between the aromatic residues of Y99 and F174, suggesting the presence of extensive hydrophobic interactions in the S4 pocket of the **2r**-FXa complex. Indeed, the cyclopropane carbons coincide closely with a high-affinity hydrophobic region predicted by the GRID program (Fig. 4b). ¹⁶

In addition, part of the side-chain CH₂-*N*-pyrrolidinyl group is close to the backbone of E97 and T98. Although the precise origin is unclear, the increase in potency from this pendant cyclopropyl methylene substituents is likely due to a combination of (1) the favorable hydrophobic interactions between the methyl pyrrolidinyl group with the edges of F174 and Y99, and the backbone atoms of E97 and T98, and (2) the conformational bias toward the perpendicular conformation of the phenyl cyclopropane placing the bulky methylene side-chain orthogonal to the plane of the phenyl ring. Little in the X-ray models suggests that H-bonding or Coulombic interactions with the 90's loop contribute to potency. Neither can

^b K_i values were obtained from purified human enzymes and were averaged from multiple determinations $(n \ge 2)$. K_i and PT values were measured as described in Ref. 5a.

a strong case be made for the interactions of the cationic groups with the aryl rings of Y99 or F174 in a π -cation sense, given the concentration of the charge at the periphery of the S4 pocket.¹⁷

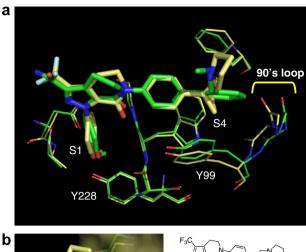
In Table 2, the in vitro FXa activity of compounds bearing *alpha*-substituted phenylcyclopropyl P4 groups is compared with corresponding compounds containing the biphenyl P4 moieties. In general, the phenylcyclopropyl analogs were more potent than the corresponding *ortho*-substituted biphenyl P4 analogs, for example, **2n**, **2o**, and **2s** showed 20- to 30-fold improvement of FXa binding affinity compared with **1a**, **1c**, and **1d**, respectively. In addition, compounds with phenylcyclopropyl P4 moieties were threefold to fivefold more potent in anticoagulant activity as measured by the EC_{2x} in the prothrombin assay.

Comparison of the crystal structures of the two series may offer some insight into the basis for the higher affinity of the phenylcyclopropane series relative to the biphenyl analogs. Among the

Table 2 a

Compound	P4	FXa ^a K _i (nM)	PT ^a EC _{2x} (μM)
2 m	-H	0.18	1.8
1b	- -\	0.28	8
2n	- -\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	0.035	1.3
1a	- -\\n_	0.41	6.4
20	-H	0.042	2.3
1c	HN(1.4	nd
2s	-I-	0.038	2.3
1d	HQ N	0.76	8.2

^a K_i 's obtained from purified human enzymes and were averaged from multiple determinations ($n \ge 2$). K_i and PT values are measured according to Ref. 5a.



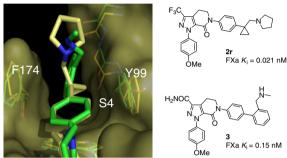


Figure 5. (a) Superposition of the X-ray structures of **3**-FXa (in green) bearing an *ortho*-substituted biphenyl P4 group and **2r**-FXa (in light yellow). Graphics were created using PyMol.¹⁵ (b) View of the P4 region in the above overlay of the X-ray structures of **3**-FXa (in green) and **2r**-FXa (in light yellow). The transparent surface of S4 from **2r**-FXa is shown in light yellow. Graphics were created using PyMol.¹⁵

fused pyrazole analogs for which we have crystal structures, compound 3^{3a} was structurally similar to the phenylcyclopropyl analog **2r**, the principle difference being the presence of an amide group at the C3 position in compound 3. Figure 5a shows the superposition of the crystal structures for the 2r- and 3-FXa complexes. In the 2r-FXa complex, the side-chain of Y99 has moved 0.48-0.78 Å to accommodate the wider cyclopropane group. This is consistent with the literature observations¹⁸ that the side-chains of Y99 and F174 can move to accommodate inhibitors of different sizes. Perhaps more striking is an observed shift (\sim 1Å) between the 97–99 backbones in the two complexes, resulting from an expansion of the S4 pocket in the **3**-FXa complex to accommodate the biphenyl group. This shift has been consistently observed with other inhibitor-FXa complexes involving biphenyl P4 groups in the pyrazole bicyclic series.3 The marked shift of the 97-99 backbone in the biphenyl analogs may involve some degree of strain and may reduce favorable interactions between the 97-99 backbone and N-alkyl side-chain as observed in the cyclopropane analog 2r-FXa complex (Fig. 4), and consequently may contribute to the observed decrease in the binding affinity of the biphenyl analogs. While the electron density of these side-chains is generally not very well defined in the crystal structure, the closer proximity of the alkyl sidechain to the interior of the S4 pocket as suggested by Figure 5, may result in more favorable binding interactions in the phenylcyclopropane series.

Because of the low anticoagulant activity of the *ortho*-substituted biphenyl P4 analogs **1a–1d** with a CF₃ C3 group (Table 2, PT EC_{2x} > 6 μ M), these compounds were not selected for dog PK studies. Table 3 compares the in vitro and pharmacokinetic profile of a representative phenylcyclopropyl analog **2n** with that of razaxaban. ^{5a} Compound **2n** was tenfold more potent in binding affinity (FXa K_1) than razaxaban, but only twofold more potent in

Table 3 Comparison of the in vitro and in vivo profile of a representative compound $2n^a$ with that of razaxaban

	F _S C NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN	Razaxaban
FXa K _i (nM)	0.035	0.19
PT EC _{2x} (μM)	1.3	3.0
Thrombin K_i (nM)	191	540
Trypsin K _i (nM)	>15,000	>10,000
Caco-2 $P_{\rm app} \times 10^{-6} ({\rm cm/s})$	8.5	5.6
HLM $t_{1/2}$ (min)	110	36
Cl (L/Kg/h) ^b	1.2	1.1
V_{dss} (L/Kg)	15	5.3
$pot_{1/2} (h)^c$	12	3.4
F%b	53	84

 $^{\rm a}$ Compound ${\bf 2n}$ was dosed as TFA salt in an N-in-1 format.

b Iv dose: 0.5 mg/kg. c Po dose: 0.2 mg/kg.

anticoagulant activity (PT EC_{2x}). Thus, it is likely that ${\bf 2n}$ is highly protein bound. Both ${\bf 2n}$ and razaxaban were selective against other related serine proteases (only trypsin and thrombin are illustrated). Both compounds show similarly moderate cell permeability. Compound ${\bf 2n}$ has better human liver microsomal stability, moderate clearance, large $V_{\rm dss}$, and good oral availability in dogs similar to razaxaban. Compound ${\bf 2n}$ has a longer $t_{1/2}$ because of the larger $V_{\rm dss}$.

Scheme 1 illustrates the synthesis of fused pyrazole analogs bearing α-substituted phenylcyclopropyl P4 groups exemplified by the preparation of **2c**, **2d**, **2l**, **2r**, and **2w**. Treatment of δ -valerolactam with phosphorus pentachloride in hot chloroform yielded 3,3-dichloro-2-piperidinone. Using lithium carbonate as the base, a good yield of 3-chloro-5,6-dihydro-2(1H)-pyridinone 4 was obtained by heating the above dichloride in DMF. The pyridinone 4 was then coupled with the readily available 2,2,2-trifluoro-N'-(4methoxyphenyl)acetohydrazonoyl bromide 5¹⁹ in the presence of triethylamine in EtOAc at reflux. After simple silica gel chromatography purification, the desired pyrazole bicyclic core 6 was obtained in a moderate 39% yield. On the other hand, electrophilic aromatic iodination of 1-phenylcyclopropyl carboxylic acid with sodium iodate and a catalytic amount of sulfuric acid in hot acetic acid afforded the 4-iodophenyl analog 7 regioselectively. The molecular skeleton 8 needed for P4 SAR studies was assembled from the pyridinone fragment 6 and the aryl iodide fragment 7 via amidation of the aryl iodide in the presence of Copper(I) iodide using 1,10-phenanthroline as the ligand and potassium carbonate as the base. The acid group in 8 was transformed to the alcohol in 2c via reduction of the mixed anhydride with sodium borohydride. A variety of final compounds were then obtained through manipulation of 2c. For example, mesylation of 2c, azide displacement followed by reduction of the azide provided the methylamine analog 21. Treatment of 2c with pyridinium chlorochromate and sodium acetate gave the aldehyde, which was subjected to reductive amination with pyrrolidine to afford the pyrrolidinylmethyl analog **2r**. Alcohol **2c** was transformed to the bromide with carbon tetrabromide and triphenylphosphine. The resulting bromide was then displaced with 2-methylimidazole to generate the imidazole analog 2w. The methylether 2d was prepared via methylation of the alcohol 9 with trimethyloxonium tetrafluoroborate, followed by amidation of the resulting iodide **10** with pyridinone **6**.

We identified *alpha*-substituted phenylcyclopropanes as viable conformational surrogates of *ortho*-substituted biphenyl groups,

Scheme 1. Reagents and conditions: (a) PCl_5 (3 equiv), CCl_3 , CCl_4 , CCl_5 , $CCl_$

generating a series of extremely potent pyrazole bicyclic-based FXa inhibitors bearing alpha-substituted phenylcyclopropyl P4 groups. These compounds exhibited FXa K_i 's in the pM range, had highly potent anticoagulant activity in human plasma, and were generally more active than the corresponding ortho-substituted biphenyl counterparts. Structural modifications of the alphasubstituted phenylcyclopropane P4 analogs to modulate in vitro and in vivo properties shall be reported in future communications. X-ray structure analysis of **2r**-FXa revealed that the cyclopropyl ring adopted a perpendicular conformation, as designed. Comparison of the X-ray structures of the phenylcyclopropyl and the orthobiaryl P4 analogs suggests that the increase in FXa affinity observed with the phenylcyclopropyl analogs may result from one or more of the following: (1) the extensive hydrophobic interactions in the S4 pocket of the phenylcyclopropyl analogs; (2) the slightly reduced strain energy involved in the phenylcyclopropyl analogs as a result of the conformational bias to adopt the preferred perpendicular conformation; (3) the potential strain generated by the 1 Å shift of the backbone of the 90's loop in the biphenyl analogs. Because of the broad application of the *ortho*-substituted biphenyl moieties, use of the perpendicular conformation of the *alpha*-substituted arylcyclopropyls to mimic the bioactive conformation of *ortho*-substituted biaryl moieties can be generalized and applied to achieve structural diversification and novelty in future drug design.

Acknowledgments

The authors thank Dr. Chubiao Xue for insightful personal discussions and Dr. Steven Sheriff for depositing **2r**-FXa complex into PDB.

References and notes

- For recent review papers of FXa inhibitors, see: (a) Quan, M.; Smallheer, J. Curr. Opin. Drug Discov. Dev. 2004, 7, 460; (b) Walenga, J. M.; Jeske, W. P.; Hoppensteadt, D.; Fareed, J. Curr. Opin. Invest. Drugs 2003, 4, 272; (c) Gould, W. R.; Leadley, R. J. Curr. Pharm. Design 2003, 9, 2337; (d) Quan, M. L.; Wexler, R. R. Curr. Topics Med. Chem. 2001, 1, 137 (Hilversum, Netherlands); (e) Zhu, B.; Scarborough, R. M. Ann. Rpts. Med. Chem. 2000, 35, 83.
 (a) Lassen, M. R.; Davidson, B. L.; Gallus, A.; Pineo, G.; Ansell, J.; Deitchman, D.
- (a) Lassen, M. R.; Davidson, B. L.; Gallus, A.; Pineo, G.; Ansell, J.; Deitchman, D. Blood 2003, 102, 11; (b) Straub, A.; Pohlmann, J.; Lampe, T.; Pernerstorfer, J.; Schlemmer, K-H.; Reinemer, P.; Perzborn, E.; Roehrig, S. J. Med. Chem. 2005, 48, 5900; (c) Eriksson, B. L.; Borris, L.; Dahl, O. E.; Haas, S.; Huisman, M. V.; Kakkar, A. K. J. Thromb. Haemost. 2006, 4, 121; (d) Hampton, T. JAMA 2006, 295, 743; (e) Wong, P. C.; Crain, E. J.; Watson, C. A.; Xin, B.; Wexler, R. R.; Lam, P. S. Y.; Pinto, D. J.; Luettgen, J. M.; Knabb, R. M. Thromb. Haemost. 2008. doi:10.1111/j.1538-7836.2008.02939.x; (f) Turpie, A. G. G. Arterioscler. Thromb. Vasc. Biol. 2007, 27, 1238. and references therein.
- 3. (a) Pinto, D. J. P.; Orwat, M. J.; Rossi, K. A.; Alexander, R. S.; Smallwood, A.; Wong, P. C.; Rendina, A.; Luettgen, J. M. J.; Knabb, R. M.; He, K.; Xin, B.; Wexler, R. R.; Lam, P. Y. J. Med. Chem. 2007, 50, 5339; (b) Pinto, D. J. P.; Orwat, M. J.; Quan, M. L.; Han, Q.; Galemmo, R. A.; Amparo, E.; Wells, B.; Ellis, C.; He, M. Y.; Alexander, R. S.; Rossi, K. A.; Smallwood, A.; Wong, P. C.; Luettgen, J. M.; Rendina, A. R.; Knabb, R. M.; Mersinger, L.; Kettner, C.; Bai, S.; He, K.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2006, 16, 4141; (c) Pinto, D. J. P.; Galemmo, R. A.; Quan, M. L.; Orwat, M. J.; Clark, C.; Li, R.; Wells, B.; Woerner, F. Alexander, R. S.; Rossi, K. A.; Smallwood, A.; Wong, P. C.; Luettgen, J. M.; Rendina, A. R.; Knabb, R. M.; He, K.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2006, 16, 5584; (d) Li, Y-L.; Fevig, J. M.; Cacciola, J.; Buriak, J.; Rossi, K. A.; Jona, J.; Knabb, R. M.; Luettgen, J. M.; Wong, P. C.; Bai, S. A.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2006, 16, 5176; (e) Fevig, J. M.; Cacciola, J.; Buriak, J.; Rossi, K. A.; Knabb, R. M.; Luettgen, J. M.; Wong, P. C.; Bai, S. A.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2006, 16, 5176; (e) Fevig, J. M.; Cacciola, J.; Buriak, J.; Rossi, K. A.; Knabb, R. M.; Luettgen, J. M.; Wong, P. C.; Bai, S. A.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2006, 16, 3755.
- Qiao, J. X.; Cheng, X.; Smallheer, J. M.; Galemmo, R. A.; Spencer, D.; Pinto, D. J. P.; Cheney, D. L.; He, K.; Wong, P. C.; Luettgen, J. M.; Knabb, R. M.; Wexler, R. R.; Lam, P. Y. S. Bioorg. Med. Chem. Lett. 2007, 17, 1432.
- (a) Quan, M. L.; Lam, P. Y. S.; Han, Q.; Pinto, D. J.; He, M.; Li, R.; Ellis, C. D.; Clark, C. G.; Teleha, C. A.; Sun, J. H.; Alexander, R. S.; Bai, S. A.; Luettgen, J. M.; Knabb, R. M.; Wong, P. C.; Wexler, R. R. J. Med. Chem. 2005, 48, 1729; (b) Lassen, M. R.; Davidson, B. L.; Gallus, A.; Pineo, G.; Ansell, J.; Deitchman, D. Blood 2003, 102, 15a. Abstract 41.

- 6. (a) A torsional scan of 2'-methyl-biphenyl was done at MP2/cc-PVTZ//B3LYP/HF-631C*. Equilibrium geometries were obtained using Jaguar (Jaguar, version 6.5, Schrödinger, LLC, New York, NY, 2006), while subsequent single point energy calculations were done using Q-Chem v3.1. A preferred twist in the range of 40–70 degrees between the two phenyls was evident. The 90-degree torsion present in the crystal structure incurred a small amount of strain energy (1.1 kcal) which is likely to be near the margin of error.; (b) Jaguar, v6.5 Schrödinger, LLC, New York, NY, 2006.; (c) Shao, Y. et al Phys. Chem. Chem. Phys. 2006, 8, 3172.
- (a) Goodman, A. L.; Eastman, R. H. J. Am. Chem. Soc. 1964, 86, 908; (b) Closs, G. L.; Klinger, H. B. J. Am. Chem. Soc. 1965, 87, 3265; (c) Pews, R. G.; Ojha, N. D. J. Am. Chem. Soc. 1969, 91, 5769.
- (a) de Boer, J. S. A. M.; Loopstra, B. O.; Stam, C. H. Rec. Tray. Chim. Pays-Bas 1987, 106, 537; (b) Bernal, I.; Levendis, D. C.; Fuchs, R.; Reisner, G. M.; Cassidy, J. M. Struct. Chem. 1997, 8, 275.
- (a) Rademacher, P. Chem. Rev. 2003, 103, 933. and references herein; (b) Martinelli, L; Mutha, S. C.; Ketcham, R; Strait, L. A.; Cavestri, R. J. Org. Chem. 1972, 37, 2278; (c) Van-Catledge, F. A. J. Am. Chem. Soc. 1973, 95, 1173; (d) Sorriso, S.; Stefani, F.; Semprini, E.; Flamini, A. J. Chem. Soc., Perkin Trans. 2: Phy. Org. Chem. 1976, 4, 374; (e) Parr, W. J. E.; Schaefer, T. J. Am. Chem. Soc. 1977, 99, 1033; (f) Castan, P.; Lopez, A.; Martino, R. Tetrahedron 1979, 35, 1093; (g) Drumright, R. E.; Mas, R. H.; Merola, J. S.; Tanko, J. M. J. Org. Chem. 1990, 55, 4098; (h) Shen, Q.; Wells, C.; Traetteberg, M.; Bohn, R. K.; Willis, A.; Knee, J. J. Org. Chem. 2001, 66, 5840.
- (a) Prins, I.; Verhoeven, J. W.; DeBoer, Th. J.; Worrell, C. Tetrahedron 1977, 33, 127; (b) Fischer, P.; Kurtz, W.; Effenberger, F. Chem. Ber. 1973, 106, 549.
- Researchers from BI reported a dual FXa/thrombin inhibitor containing a benzimidazolyl cyclopropyl P4 group with an π-character alpha substituent (PDB entry: 1G2L) adopts a bisected conformation. Nar, H.; Bauer, M.; Schmid, A.; Stassen, J.-M.; Wienen, W.; Priepke, H. W. M.; Kauffmann, I. K.; Ries, U. J.; Hauel, N. H. Structure 2001, 9, 29.
- (a) Torsional scans of cyclopropylbenzene and alpha-methyl phenylcyclopropane were done at MP2/6311C*//B3LYP/HF-631C*. Equilibrium geometries were obtained using Jaguar (Jaguar, version 6.5, Schrödinger, LLC, New York, NY, 2006), while subsequent single point energy calculations were done using Q-Chem v3.01.; (b) Cheney, D. L.; Qiao, J. X. Abstract of papers, 233rd ACS National Meeting, Chicago, IL, March 25-29, 2007, MEDI-198.
- 13. Experimental pKa of 2-amino-thiazole = 5.6 at 25 °C (a) Gabryszewski, G.; Kulig, J.; Lenarcik, B. *Polish J. Chem.* **1982**, 56, 55; Experimental pKa of 1,2 dimethylimidazole = 8.2 at 25 °C (b) Lenarcik, B.; Barszcz, B. *J. Chem. Soc. Dalton* **1980**, 24.
- 14. PDB Deposition number for 2r-FXa complex is 3CS7.
- DeLano, W. L. The PyMOL Molecular Graphics System (2002) on World Wide Web http://www.pymol.org.
- The program GRID is a forcefield-based method for contouring various types of intermolecular interactions over protein surfaces. Molecular Discovery, Discovery Via, Stoppani, Ponte San Giovanni, Italy.
- Schaerer, K.; Morgenthaler, M.; Paulini, R.; Obst-Sander, U.; Banner, D. W.; Schlatter, D.; Benz, J.; Stihle, M.; Diederich, F. Angew. Chem. Int. Ed. 2005, 44, 4400.
- (a) Adler, M.; Kochanny, M. J.; Ye, B.; Rumennik, G.; Light, D. R.; Biancalana, S.; Whitlow, M. Biochemistry 2002, 41, 15514; (b) Maignan, S.; Guilloteau, J. P.; Pouzieux, S.; Choi-Sledeski, Y. M.; Becker, M. R.; Klein, S. I.; Ewing, W. R.; Pauls, H. W.; Spada, A. P.; Mikol, V. J. Med. Chem. 2000, 43, 3226; (c) Adler, M.; Davey, D. D.; Phillips, G. B.; Kim, S.-H.; Jancarik, J.; Rumennik, G.; Light, D. R.; Whitlow, M. Biochemistry 2000, 39, 12534.
- 19. Tanaka, K.; Maeno, S.; Mitsuhashi, K. *Chem. Lett.* **1982**, *4*, 543.